

STEREOCHEMISTRY OF THE ESTER SIDE CHAIN OF THE GERMACRANOLIDES OF *VIGUIERA HYPARGYREA*¹

GUILLERMO DELGADO,* LAURA ALVAREZ, MANUEL SORIANO-GARCÍA,
RUBÉN A. TOSCANO, RACHEL MATA,² and ROGELIO PEREDA-MIRANDA³

Instituto de Química de la Universidad Nacional Autónoma de México, Ciudad Universitaria,
Circuito Exterior, Coyoacán 04510, México, D.F.

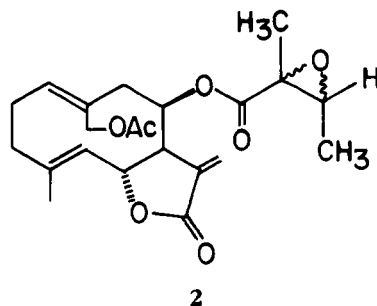
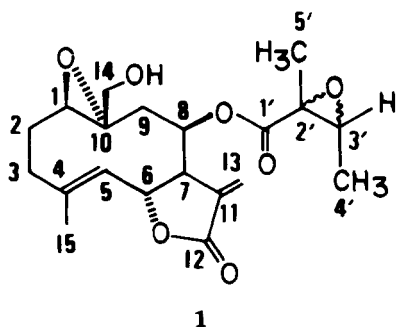
In earlier articles (1-4), the chemical constituents of several *Viguiera* species (Compositae) were reported, and the results obtained indicate that the terpenoids are useful in discerning infrageneric groups and are in agreement with the accepted biogenetic relationships between members of the Helianthineae (*Helianthus*, *Tithonia*, *Syncretocarpus*, among others) (5). Furthermore, the presence of sesquiterpene lactones of advanced biogenetic complexity in *Helianthus* and *Tithonia* (but not in *Viguiera*) supports the putative derivatization of these genera from a Viguieroid stock (6).

In accordance with these findings, several novel germacranolides have recently been isolated from *Viguiera hypargyrea* (6). However, the complete

stereochemical expression of **1** and **2** were not established, because the configuration of the five carbon ester side chain was not determined.

The two possible diastereoisomers (2'*R*,3'*R* and 2'*S*,3'*S*) of epoxyangeloyloxy sesquiterpene lactones are found in nature (7), but arguments based on spectral data are not reliable guides for discrimination. Therefore, an X-ray structure determination of **1** or **2** (or a derivative) was desirable. The natural products did not give suitable crystals, but acid treatment of **3** [obtained by acetylation of **1** or by epoxidation of **2** (6)] with HCl afforded the crystalline chlorhydrin **4** (see Experimental) which was satisfactory for this study.

Details of this analysis are given in the experimental section and listings of per-



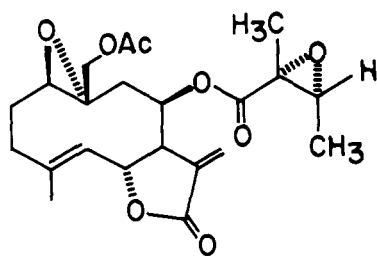
¹Contribution No. 845 of Instituto de Química de la UNAM.

²Present address: Departamento de Química Farmacéutica y Productos Naturales, División de Estudios de Posgrado de la Facultad de Química, Universidad Nacional Autónoma de México, Circuito Escolar, Ciudad Universitaria, Coyoacán 04510, México, D.F.

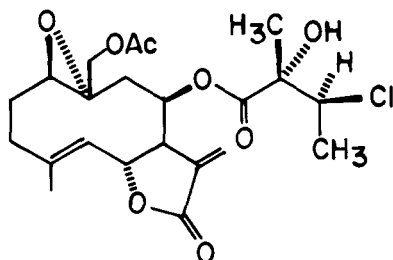
³Present address: Departamento de Química Orgánica de la Escuela Nacional de Ciencias Biológicas, Instituto Politécnico Nacional, México 11340, D.F.

tinent crystallographic data are available as supplementary material. Atomic coordinates for chlorhydrin **4** are given in Table 1, and Figure 1 is a computer-generated drawing of the enantiomer which, based on the anomalous scattering of the chlorine atom, also represents the absolute configuration.

The 2'*R*,3'*S* configuration of the ester side chain of **4** (Figure 1) indicates a 2'*R*,3'*R* stereochemistry of the



3



4

epoxyangeloyl residue of **3** (and, therefore, of **1** and **2**) due the regio-differentiated S_N2 ring opening of the oxirane with inversion at C-3'.

The $^1D^{14}, ^{15}D_5$ crown conformation of the macrocycle depicted in Figure 1 is not significantly different from those determined by X-ray analysis for other germacranolides with *trans*-lactones closed

to C-6 such as hanphyllin (**8**) and euserotin (**9**), although the four extreme conformers (due to diastereofacial arrangements of the double bonds of their equivalents) are found in the germacrolide series (**10**).

EXPERIMENTAL

8 β -[Epoxyangeloyloxy]-14-hydroxy-tithifolin

TABLE 1. Positional Parameters ($\times 10^4$) and Equivalent Isotropic Temperature Factors (\AA^2) for Compound **4**

	x	y	z	U_{eg}
C1	3512(2)	7418(2)	6708(2)	70(1)
O(1)	10465(4)	2184(4)	10453(3)	49(1)
O(2)	1833(4)	3608(4)	12816(3)	49(1)
O(3)	5411(4)	4098(3)	8923(3)	36(1)
O(4)	-306(4)	5776(5)	12540(4)	77(2)
O(5)	9051(4)	1375(4)	8249(3)	48(1)
O(6)	5888(4)	4230(4)	5293(3)	57(1)
O(7)	7413(4)	5407(4)	6844(3)	54(1)
O(8)	8222(6)	-1220(5)	8262(4)	89(2)
C(1)	8936(5)	1924(5)	11753(4)	44(2)
C(2)	8759(6)	115(6)	12729(4)	51(2)
C(3)	6878(6)	-78(6)	13889(4)	53(2)
C(4)	5345(6)	478(5)	13216(4)	43(2)
C(5)	4764(5)	2163(5)	13029(4)	38(2)
C(6)	3753(5)	3141(5)	12020(4)	37(2)
C(7)	4597(5)	4918(5)	11160(4)	39(2)
C(8)	6136(5)	4953(5)	9739(4)	37(2)
C(9)	8113(5)	4311(5)	9750(4)	39(2)
C(10)	8586(5)	2411(5)	10360(4)	37(2)
C(11)	2913(6)	6089(5)	11053(4)	46(2)
C(12)	1276(6)	5226(6)	12172(5)	51(2)
C(13)	2789(8)	7606(7)	10180(5)	71(2)
C(14)	8135(6)	1030(5)	9784(4)	41(2)
C(15)	4758(7)	-907(6)	12688(5)	60(2)
C(16)	6074(5)	4570(5)	7499(4)	37(1)
C(17)	4885(5)	3967(5)	6779(4)	40(2)
C(18)	3058(6)	5126(6)	7010(4)	49(2)
C(19)	4449(7)	2050(6)	7369(4)	54(2)
C(20)	1955(7)	4994(8)	6061(6)	72(2)
C(21)	8992(6)	94(5)	7610(4)	43(2)
C(22)	9882(7)	603(6)	6042(4)	59(2)

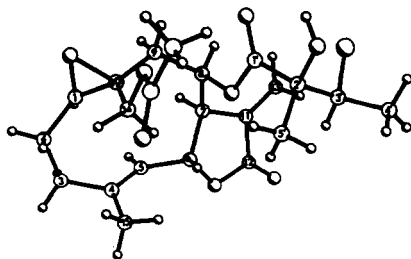


FIGURE 1. Computer perspective drawing of 4

[1] and 8 β -[epoxyangeloyl]-14-acetoxy-epupatolide [2] were the substrates for 3 (via acetylation of 1 or epoxidation of 2) as described in the previous work (6).

ACID TREATMENT OF 3 TO 4.—8 β -[Epoxyangeloyloxy]-14-acetoxy-tithifolin [3] (80 mg) was treated with dry HCl gas in Et₂O (75 ml) at -5° for 2 min. The reaction mixture was diluted with H₂O, washed with saturated aqueous NaHCO₃, and the organic phase dried and evaporated. The residue was chromatographed over Si gel column using hexane-EtOAc (3:1) as eluent. The product (43 mg) was crystallized from Me₂CO/*i*Pr₂O to afford crystals of 4, mp 220–222 $^{\circ}$; ir (CHCl₃) 3560, 1765, 1740, 1735, 1640 cm⁻¹; ¹H nmr (80 MHz, CDCl₃) δ 6.32 (1H, d, *J*=3.5 Hz, H-13), 5.72 (1H, m, H-8), 5.62 (1H, d, *J*=3.5 Hz, H-13'), 5.37 (1H, dq, *J*=10, 1 Hz, H-5), 5.12 (1H, dd, *J*=9, 10 Hz, H-6), 4.16 (1H, d, *J*=9 Hz, H-14a), 4.01 (1H, d, *J*=9 Hz, H-14b), 3.25 (1H, m, H-9a), 2.93 (1H, q, *J*=7 Hz, H-3'), 2.05 (3H, s, CH₃CO), 1.84 (3H, d, *J*=1 Hz, 15CH₃), 1.48 (3H, *J*=7 Hz, 4'CH₃), 1.31 (3H, s, 5'CH₃); ¹³C nmr (20 MHz, CDCl₃) δ 173.32 (s, C-1'), 170.69 (s, CH₃CO), 168.77 (s, C-12), 144.57 (s, C-4), 135.41 (s, C-11), 124.93 (d, C-5), 122.21 (t, C-13), 77.34 (d, C-3'), 73.75 (d, C-6), 69.93 (d, C-2'), 66.88 (d, C-1), 64.85 (t, C-14), 62.69 (d, C-8), 59.85 (s, C-10), 53.21 (d, C-7), 38.23 (t, C-9), 35.80 (t, C-3), 24.53 (q, C-5'), 22.11 (t, C-2), 20.81 (q, CH₃CO), 18.19 (q, C-4'), 17.18 (q, C-15); cims *m/z* (rel. int.) 459 (9) [M+1]⁺, 457 (29), 439 (15), 417 (34), 415 (92), 381 (10), 379 (32), 305 (33), 263 (42), 246 (18), 245 (75), 227 (100), 107 (5). (Found: C, 58.18%; H, 6.44. C₂₂H₂₉O₈Cl requires: C, 57.90; H, 6.30).

X-RAY STRUCTURE ANALYSIS.—The chlorhydrin 4 was crystallized by slow evaporation from EtOAc solution. Crystal data: C₂₂H₂₉O₈Cl, *M*_w=456.9, triclinic, space group P1, *a*=7.689 (4), *b*=7.869 (4), *c*=10.189 (4) Å, α =75.42 (4), β =70.27 (3), γ =82.19 (4) $^{\circ}$, *V*=561 (1) Å³, *D*_c=1.35 g cm⁻³, *F*(000)=242, *T*=293 K, *Z*=1, μ (MoK α)=2.11 cm⁻¹. Using a crystal 0.18 \times 0.27 \times 0.32 mm, lattice parameters were obtained from 25 machine-centered re-

flections with $5.3 < 2\theta < 21.4^{\circ}$. Reflections in a hemisphere of reciprocal space were measured with an index range of *h* 0/7, *k* \pm 8, *l* \pm 10, using ω scanning mode with graphite-monochromatized MoK α radiation on a Nicolet R3m four circle diffractometer, variable scan speed (max 29.3 $^{\circ}$ /min; min 3.91 $^{\circ}$ /min), scan width 1.0 ($^{\circ}$), two standard reflections (1,1,-1; 0,1,-2) monitored every 50 measurements. The intensities were corrected for Lorentz and polarization effects, but no absorption corrections were applied. Of the 1595 reflections within the 2θ range of $3 - 45^{\circ}$ collected, 1457 had values of $3\sigma(I)$ their estimated standard deviations, and these were used in the final refinement of structural parameters. The structure was solved by direct methods (11), using 200 phases with $|E| > 1.2$ and 12 reflections in the starting set, and refined by cascade matrix least squares techniques with anisotropic temperature factors for the non hydrogen atoms. The H atoms of the tertiary CH, secondary CH₂, and primary CH₃ groups were assigned coordinates based on the expected bonding geometry. All H atoms coordinates were refined with a fixed anisotropic temperature factor of *U*=0.06 Å². The function minimized was $\sum \omega(\Delta F)^2$ with a statistical weight of the form $\omega = \sigma^2(F_o) + 0.0005(F_o)^2$, where σ is the standard deviation of the observed amplitudes based on counting statistics. The conventional R factor was 0.035 and $\omega R = 0.040$ ($\omega R = |\sum \omega(|F_o| - |F_c|)^2 / \sum |F_o|^2|^{1/2}$, scattering factors for Cl, O, C, and H atoms were from International Tables for X-ray Crystallography (12), and the isotropic extinction parameter $x = 0.00173$. A final difference Fourier synthesis revealed no peaks higher than those assigned earlier to hydrogen atoms, and the weighting appeared reasonable.

As all correction terms allowing for anomalous scattering were introduced at the beginning of refinements, a significance test (13) was used to discriminate between the two possible enantiomorphs. For comparison with *R*_g+ = 0.0400, the *R*- of the other enantiomorph was obtained by a least-squares refinement calculated with inverted coordinates. The residual for this enantiomorph was *R*_g- = 0.0402, *R*-/*R*+ Hamilton's *R*-factor test showed that the second absolute

configuration can be rejected at a confidence level >99.0%.

Although this seemed quite satisfactory, a new Hamilton's R-factor test using data collected with Cu-filtered radiation was performed, confirming the last assignment as a high confidence level (>99.5%), $R+ = .0957$, $R- = .1045$, $R-/R+ = 1.092$.

The final atomic coordinates for the nonhydrogen atoms are listed in Table 1, and pertinent data are available as supplementary material.⁴

ACKNOWLEDGMENTS

This work was supported in part by the Consejo Nacional de Ciencia y Tecnología, CONACYT, México.

LITERATURE CITED

1. G. Delgado, L. Alvarez, and A. Romo de Vivar, *Phytochemistry*, **23**, 675 (1984).
2. G. Delgado, L. Alvarez, and A. Romo de Vivar, *Phytochemistry*, **23**, 2674 (1984).
3. G. Delgado, H. Cárdenas, G. Peláez, A. Romo de Vivar, and R. Pereda-Miranda, *J. Nat. Prod.*, **47**, 1042 (1984).
4. G. Delgado, L. Alvarez, and A. Romo de Vivar, *Phytochemistry*, **24**, 2736 (1985).
5. A. Romo de Vivar and G. Delgado, *Bol. Soc. Chil. Quím.*, **30**, 79 (1985).
6. L. Alvarez, R. Mata, G. Delgado, and A. Romo de Vivar, *Phytochemistry*, **24**, 2973 (1985).
7. W. Herz and N. Kumar, *Phytochemistry*, **20**, 1339 (1981).
8. R. Mata, G. Delgado, and A. Romo de Vivar, *Phytochemistry*, **24**, 1515 (1985).
9. W. Herz, R. DeGroote, R. Murari, N. Kumar, and J.F. Blount, *J. Org. Chem.*, **44**, 2784 (1979).
10. K. Tori, I. Horibe, Y. Tamura, K. Kuriyama, H. Tada, and K. Takeda, *Tetrahedron Lett.*, 387 (1976).
11. G.M. Sheldrick, "SHELXTL' revision 3: An Integrated System for Solving, Refining and Displaying Crystal Structures from Diffraction Data," University of Göttingen, Federal Republic of Germany, 1981.
12. "International Tables for X-ray Crystallography," vol. 4, Kynoch Press, Birmingham, England, 1974.
13. W.C. Hamilton, *Acta Cryst.*, **18**, 502 (1965).

Received 6 June 1986

⁴The supplementary material is available on request to the senior author. Atomic coordinates for the structure have been deposited with the Cambridge Crystallographic Data Centre and can be obtained on request from Dr. Olga Kennard, University Chemical Laboratory, Lensfield Rd., Cambridge, CB2 1EW, UK.